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Type I interferons and microbial metabolites of tryptophan modulate astrocyte activity and central nervous system inflammation via the aryl hydrocarbon receptor

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Abstract

Astrocytes have important roles in the central nervous system (CNS) during health and disease. Through genome-wide analyses we detected a transcriptional response to type I interferons (IFN-Is) in astrocytes during experimental CNS autoimmunity and also in CNS lesions from patients with multiple sclerosis (MS). IFN-I signaling in astrocytes reduces inflammation and experimental autoimmune encephalomyelitis (EAE) disease scores via the ligand-activated transcription factor aryl hydrocarbon receptor (AHR) and the suppressor of cytokine signaling 2 (SOCS2). The anti-inflammatory effects of nasally administered interferon (IFN)- β are partly mediated by AHR. Dietary tryptophan is metabolized by the gut microbiota into AHR agonists that have an effect on astrocytes to limit CNS inflammation. EAE scores were increased following ampicillin treatment during the recovery phase, and CNS inflammation was reduced in antibiotic-treated mice by supplementation with the tryptophan metabolites indole, indoxyl-3-sulfate, indole-3-propionic acid and indole-3-aldehyde, or the bacterial enzyme tryptophanase. In individuals with MS, the circulating levels of AHR agonists were decreased. These findings suggest that IFN-Is produced in the CNS function in combination with metabolites derived from dietary tryptophan by the gut flora to activate AHR signaling in astrocytes and suppress CNS inflammation.

Subject terms: Autoimmunity Neuroimmunology

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References

1. Allen, N.J. *et al.* Astrocyte glypicans 4 and 6 promote formation of excitatory synapses via GluA1 AMPA receptors. *Nature* **486**, 410–414 (2012).
2. Alvarez, J.I. *et al.* The Hedgehog pathway promotes blood–brain barrier integrity and CNS immune quiescence. *Science* **334**, 1727–1731 (2011).
3. Chung, W.S. *et al.* Astrocytes mediate synapse elimination through MEGF10 and MERTK pathways. *Nature* **504**, 394–400 (2013).
4. Khakh, B.S. & Sofroniew, M.V. Diversity of astrocyte functions and phenotypes in neural circuits. *Nat. Neurosci.* **18**, 942–952 (2015).
5. Molofsky, A.V. *et al.* Astrocyte-encoded positional cues maintain sensorimotor circuit integrity. *Nature* **509**, 189–194 (2014).
6. Obermeier, B., Daneman, R. & Ransohoff, R.M. Development, maintenance and disruption of the blood–brain barrier. *Nat. Med.* **19**, 1584–1596 (2013).
7. Rieckmann, P. & Engelhardt, B. Building up the blood–brain barrier. *Nat. Med.* **9**, 828–829 (2003).
8. Sofroniew, M.V. Astrocyte barriers to neurotoxic inflammation. *Nat. Rev. Neurosci.* **16**, 249–263 (2015).
9. Tsai, H.H. *et al.* Regional astrocyte allocation regulates CNS synaptogenesis and repair. *Science* **337**, 358–362 (2012).
10. Lassmann, H. Mechanisms of white matter damage in multiple sclerosis. *Glia* **62**, 1816–1830

- (2014).
11. Mayo, L. *et al.* Regulation of astrocyte activation by glycolipids drives chronic CNS inflammation. *Nat. Med.* **20**, 1147–1156 (2014).
 12. Shao, W. *et al.* Suppression of neuroinflammation by astrocytic dopamine D2 receptors via α B-crystallin. *Nature* **494**, 90–94 (2013).
 13. Berer, K. *et al.* Commensal microbiota and myelin autoantigen cooperate to trigger autoimmune demyelination. *Nature* **479**, 538–541 (2011).
 14. Furusawa, Y. *et al.* Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* **504**, 446–450 (2013).
 15. Smith, P.M. *et al.* The microbial metabolites, short-chain fatty acids, regulate colonic T_{reg} cell homeostasis. *Science* **341**, 569–573 (2013).
 16. Baruch, K. *et al.* Aging. Aging-induced type I interferon response at the choroid plexus negatively affects brain function. *Science* **346**, 89–93 (2014).
 17. Ivashkiv, L.B. & Donlin, L.T. Regulation of type I interferon responses. *Nat. Rev. Immunol.* **14**, 36–49 (2014).
 18. Sandler, N.G. *et al.* Type I interferon responses in rhesus macaques prevent SIV infection and slow disease progression. *Nature* **511**, 601–605 (2014).
 19. Yan, Y. *et al.* CNS-specific therapy for ongoing EAE by silencing IL-17 pathway in astrocytes. *Mol. Ther.* **20**, 1338–1348 (2012).
 20. Quintana, F.J. & Sherr, D.H. Aryl hydrocarbon receptor control of adaptive immunity. *Pharmacol. Rev.* **65**, 1148–1161 (2013).
 21. Stockinger, B., Di Meglio, P., Gialitakis, M. & Duarte, J.H. The aryl hydrocarbon receptor: multitasking in the immune system. *Annu. Rev. Immunol.* **32**, 403–432 (2014).
 22. Prinz, M. *et al.* Distinct and nonredundant *in vivo* functions of IFNAR on myeloid cells limit autoimmunity in the central nervous system. *Immunity* **28**, 675–686 (2008).
 23. Fitzgerald, D.C. *et al.* Suppressive effect of IL-27 on encephalitogenic T_H17 cells and the effector phase of experimental autoimmune encephalomyelitis. *J. Immunol.* **179**, 3268–3275 (2007).
 24. Mitsdoerffer, M. & Kuchroo, V. New pieces in the puzzle: how does interferon- β really work in multiple sclerosis? *Ann. Neurol.* **65**, 487–488 (2009).
 25. Mascalfroni, I.D. *et al.* Metabolic control of type 1 regulatory T cell differentiation by AHR and HIF1- α . *Nat. Med.* **21**, 638–646 (2015).
 26. Mascalfroni, I.D. *et al.* IL-27 acts on DCs to suppress the T cell response and autoimmunity by inducing expression of the immunoregulatory molecule CD39. *Nat. Immunol.* **14**, 1054–1063 (2013).
 27. Apetoh, L. *et al.* The aryl hydrocarbon receptor interacts with c-Maf to promote the differentiation

- of type 1 regulatory T cells induced by IL-27. *Nat. Immunol.* **11**, 854–861 (2010).
28. Yeste, A., Nadeau, M., Burns, E.J., Weiner, H.L. & Quintana, F.J. Nanoparticle-mediated co-delivery of myelin antigen and a tolerogenic small molecule suppresses experimental autoimmune encephalomyelitis. *Proc. Natl. Acad. Sci. USA* **109**, 11270–11275 (2012).
29. Weidemann, A. *et al.* The glial cell response is an essential component of hypoxia-induced erythropoiesis in mice. *J. Clin. Invest.* **119**, 3373–3383 (2009).
30. Jessen, K.R. & Mirsky, R. Glial cells in the enteric nervous system contain glial fibrillary acidic protein. *Nature* **286**, 736–737 (1980).
31. Kim, R.Y. *et al.* Astrocyte CCL2 sustains immune cell infiltration in chronic experimental autoimmune encephalomyelitis. *J. Neuroimmunol.* **274**, 53–61 (2014).
32. Mc Guire, C., Prinz, M., Beyaert, R. & van Loo, G. Nuclear factor kappa B (NF- κ B) in multiple sclerosis pathology. *Trends Mol. Med.* **19**, 604–613 (2013).
33. van Loo, G. *et al.* Inhibition of transcription factor NF- κ B in the central nervous system ameliorates autoimmune encephalomyelitis in mice. *Nat. Immunol.* **7**, 954–961 (2006).
34. Boverhof, D.R. *et al.* 2,3,7,8-tetrachlorodibenzo-p-dioxin induces suppressor of cytokine signaling 2 in murine B cells. *Mol. Pharmacol.* **66**, 1662–1670 (2004).
35. Zadjali, F. *et al.* *Socs2* deletion protects against hepatic steatosis but worsens insulin resistance in high-fat-diet-fed mice. *FASEB J.* **26**, 3282–3291 (2012).
36. Axtell, R.C. *et al.* T helper type 1 and 17 cells determine efficacy of interferon- β in multiple sclerosis and experimental encephalomyelitis. *Nat. Med.* **16**, 406–412 (2010).
37. Borden, E.C. *et al.* Interferons at age 50: past, current and future impact on biomedicine. *Nat. Rev. Drug Discov.* **6**, 975–990 (2007).
38. Ross, T.M. *et al.* Intranasal administration of interferon- β bypasses the blood–brain barrier to target the central nervous system and cervical lymph nodes: a non-invasive treatment strategy for multiple sclerosis. *J. Neuroimmunol.* **151**, 66–77 (2004).
39. Yona, S. *et al.* Fate mapping reveals origins and dynamics of monocytes and tissue macrophages under homeostasis. *Immunity* **38**, 79–91 (2013).
40. Farez, M.F. *et al.* Melatonin contributes to the seasonality of multiple sclerosis relapses. *Cell* **162**, 1338–1352 (2015).
41. Gandhi, R. *et al.* Activation of the aryl hydrocarbon receptor induces human type 1 regulatory T cell-like and Foxp3⁺ regulatory T cells. *Nat. Immunol.* **11**, 846–853 (2010).
42. Li, Y. *et al.* Exogenous stimuli maintain intraepithelial lymphocytes via aryl hydrocarbon receptor activation. *Cell* **147**, 629–640 (2011).
43. Wikoff, W.R. *et al.* Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. *Proc. Natl. Acad. Sci. USA* **106**, 3698–3703 (2009).

44. Zelante, T. *et al.* Tryptophan catabolites from microbiota engage aryl hydrocarbon receptor and balance mucosal reactivity via interleukin-22. *Immunity* **39**, 372–385 (2013).
45. Schroeder, J.C. *et al.* The uremic toxin 3-indoxyl sulfate is a potent endogenous agonist for the human aryl hydrocarbon receptor. *Biochemistry* **49**, 393–400 (2010).
46. Palace, J., Leite, M.I., Nairne, A. & Vincent, A. Interferon- β treatment in neuromyelitis optica: increase in relapses and aquaporin 4 antibody titers. *Arch. Neurol.* **67**, 1016–1017 (2010).
47. Khorrooshi, R. *et al.* Induction of endogenous type I interferon within the central nervous system plays a protective role in experimental autoimmune encephalomyelitis. *Acta Neuropathol.* **130**, 107–118 (2015).
48. Dann, A. *et al.* Cytosolic RIG-I-like helicases act as negative regulators of sterile inflammation in the CNS. *Nat. Neurosci.* **15**, 98–106 (2012).
49. Ejlerskov, P. *et al.* Lack of neuronal IFN- β -IFNAR causes Lewy body- and Parkinson's disease-like dementia. *Cell* **163**, 324–339 (2015).
50. Goldmann, T. *et al.* USP18 lack in microglia causes destructive interferonopathy of the mouse brain. *EMBO J.* **34**, 1612–1629 (2015).
51. Quintana, F.J. *et al.* Control of T_{reg} and T_H17 cell differentiation by the aryl hydrocarbon receptor. *Nature* **453**, 65–71 (2008).
52. Quintana, F.J. *et al.* An endogenous aryl hydrocarbon receptor ligand acts on dendritic cells and T cells to suppress experimental autoimmune encephalomyelitis. *Proc. Natl. Acad. Sci. USA* **107**, 20768–20773 (2010).
53. Veldhoen, M. *et al.* The aryl hydrocarbon receptor links T_H17 cell-mediated autoimmunity to environmental toxins. *Nature* **453**, 106–109 (2008).
54. Bessede, A. *et al.* Aryl hydrocarbon receptor control of a disease tolerance defense pathway. *Nature* **511**, 184–190 (2014).
55. Opitz, C.A. *et al.* An endogenous tumor-promoting ligand of the human aryl hydrocarbon receptor. *Nature* **478**, 197–203 (2011).
56. Monteleone, I. *et al.* Aryl-hydrocarbon-receptor-induced signals upregulate IL-22 production and inhibit inflammation in the gastrointestinal tract. *Gastroenterology* **141**, 237–248, 248.e1 (2011).
57. Atarashi, K. *et al.* T_H17 cell induction by adhesion of microbes to intestinal epithelial cells. *Cell* **163**, 367–380 (2015).
58. Lee, Y.K., Menezes, J.S., Umesaki, Y. & Mazmanian, S.K. Proinflammatory T cell responses to gut microbiota promote experimental autoimmune encephalomyelitis. *Proc. Natl. Acad. Sci. USA* **108**, 4615–4622 (2011).
59. Ochoa-Repáraz, J. *et al.* Central nervous system demyelinating disease protection by the human commensal *Bacteroides fragilis* depends on polysaccharide A expression. *J. Immunol.* **185**, 4101–4108 (2010).

60. Viaud, S. *et al.* The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. *Science* **342**, 971–976 (2013).
61. Prinz, M., Priller, J., Sisodia, S.S. & Ransohoff, R.M. Heterogeneity of CNS myeloid cells and their roles in neurodegeneration. *Nat. Neurosci.* **14**, 1227–1235 (2011).
62. Trapnell, C. *et al.* Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks. *Nat. Protoc.* **7**, 562–578 (2012).
63. Jack, C.S. *et al.* TLR signaling tailors innate immune responses in human microglia and astrocytes. *J. Immunol.* **175**, 4320–4330 (2005).
64. Alvarez, J.I. *et al.* Focal disturbances in the blood–brain barrier are associated with formation of neuroinflammatory lesions. *Neurobiol. Dis.* **74**, 14–24 (2015).
65. Townsend, M.K. *et al.* Reproducibility of metabolomic profiles among men and women in two large cohort studies. *Clin. Chem.* **59**, 1657–1667 (2013).

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V.R., I.D.M., L.B., M.C.T., J.E.K., L.M., C.-C.C., H.K., J.I.A., M.B. and C.B.C. performed *in vitro* and *in vivo* experiments; B.P., R.Y., N.O. and N.P. performed bioinformatics analysis; N.A., G.I., C.B.C., A.P., S.J., M.P. and J.A. provided unique reagents, and discussed and/or interpreted findings; V.R. and F.J.Q. wrote the manuscript; and F.J.Q. designed and supervised the study and edited the manuscript.

Competing financial interests

The authors declare no competing financial interests.

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Supplementary information

PDF files

1. Supplementary Text and Figures (6,419 KB)
Supplementary Figures 1–7 and Supplementary Table 1–4

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